

# Gut Turnover: An Evolutionary Adaptation to Mouthbrooding in

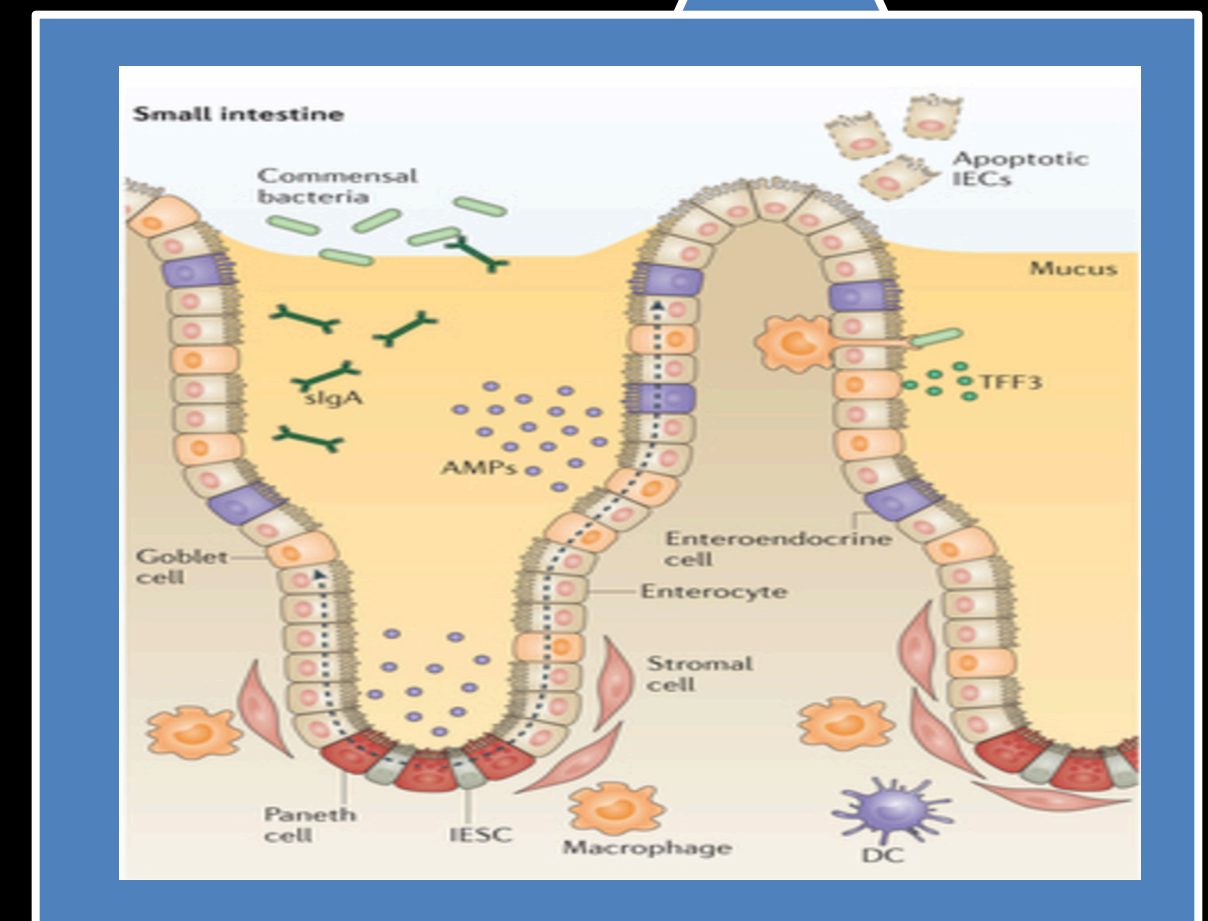
Cameron Roberts

## *Astatotilapia burtoni*

Reed College Biology 342

- Female *Astatotilapia burtoni* (mouthbrooding cichlid) voluntarily starve themselves for two weeks while their young develop.
- There are physiological mechanisms that have evolved to allow mouthbrooding
- Intestinal epithelial cell (IEC) turnover ensures integrity of the intestine,
- IEC turnover is energetically expensive
- An important tool for measuring IEC turnover is the detection of DNA fragmentation that comes from apoptotic signaling cascades (TUNEL)

If mouthbrooding females conserve energy by down regulating intestinal epithelial turnover, then there will be fewer apoptotic cells in brooding fish compared to fasted or fed females.



# TUNEL STAINING: IMMUNOHISTOCHEMISTRY

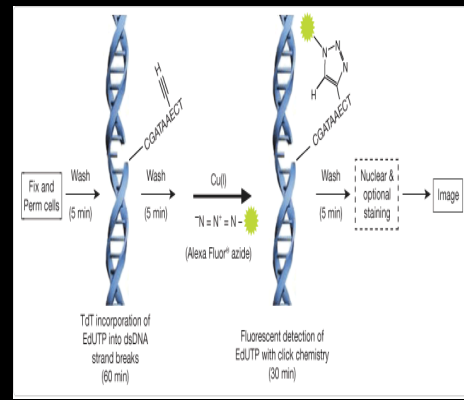
Dissection:  
brooding, starved, and fed  
female intestines



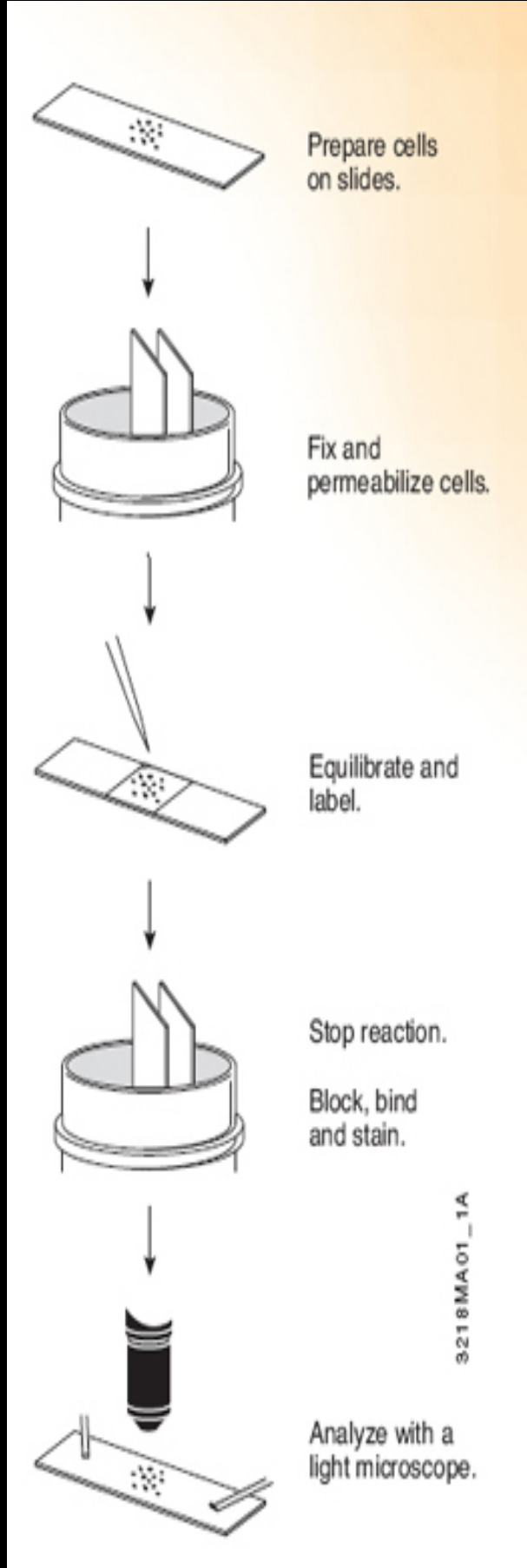
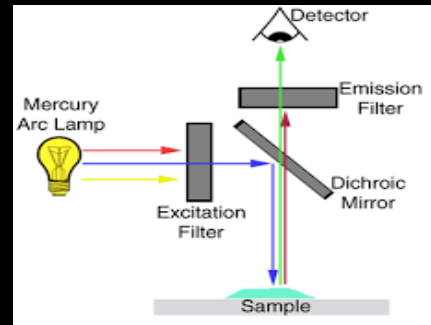
Cryostat:  
Mounted and sectioned tissue  
(17 μm slices)



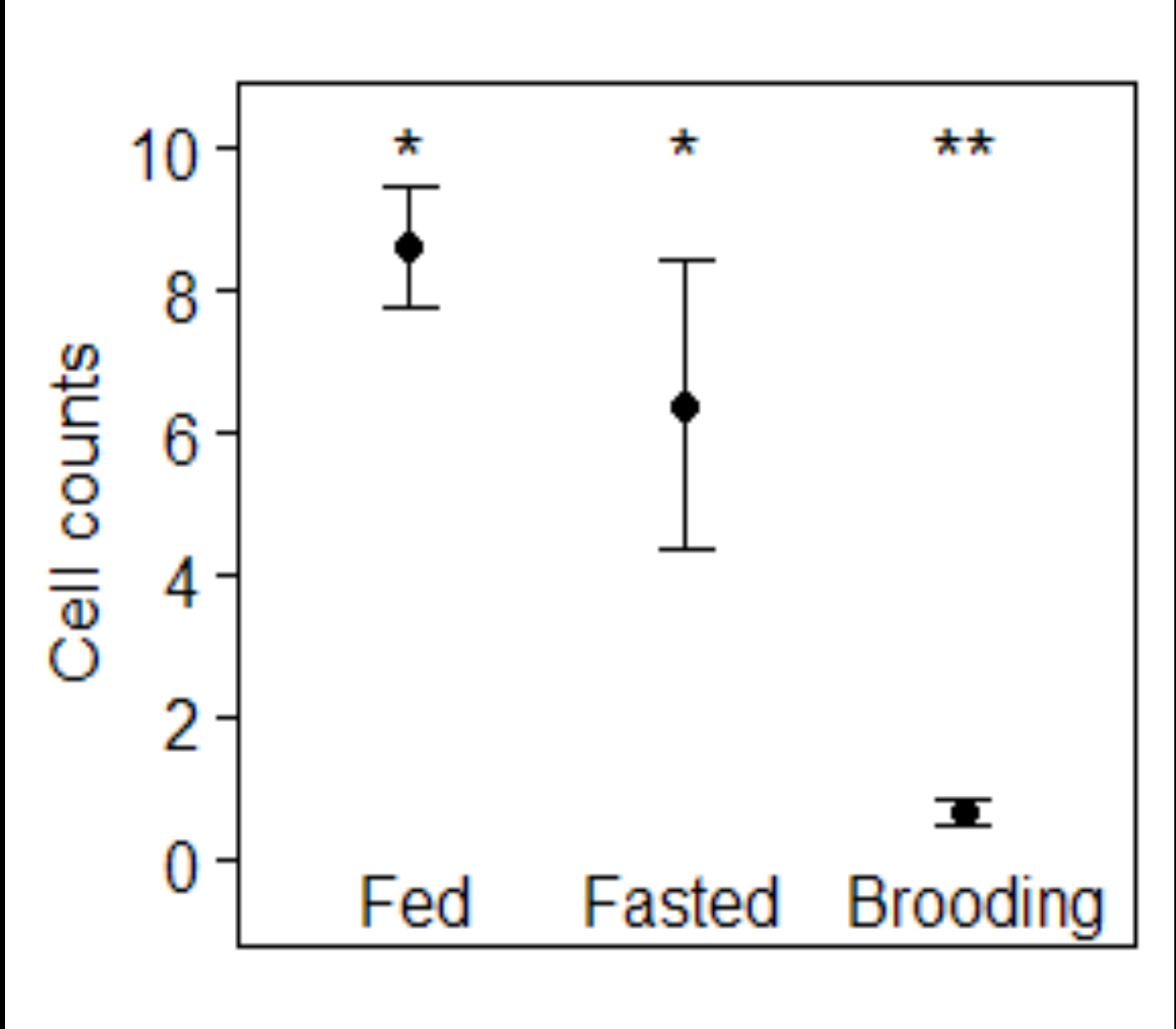
Immunohistochemistry:  
TUNEL assay detects DNA  
fragmentation by labeling the  
terminal end of nucleic acids



Analyzed using fluorescent  
microscopy

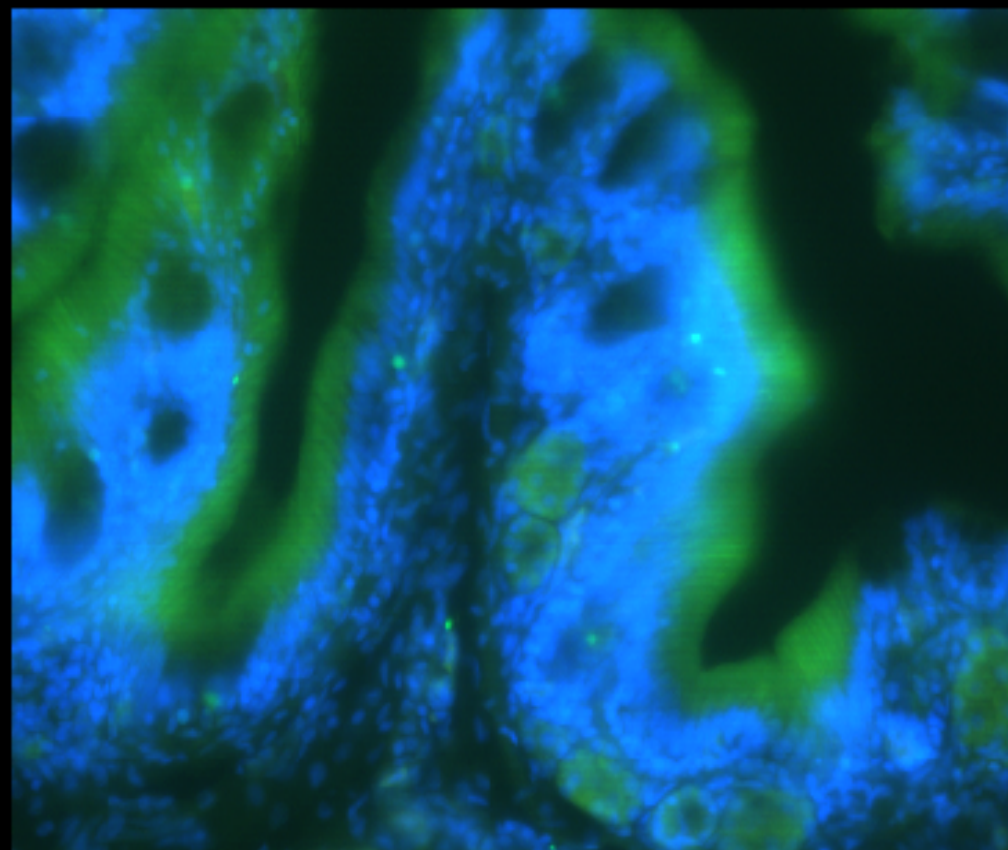
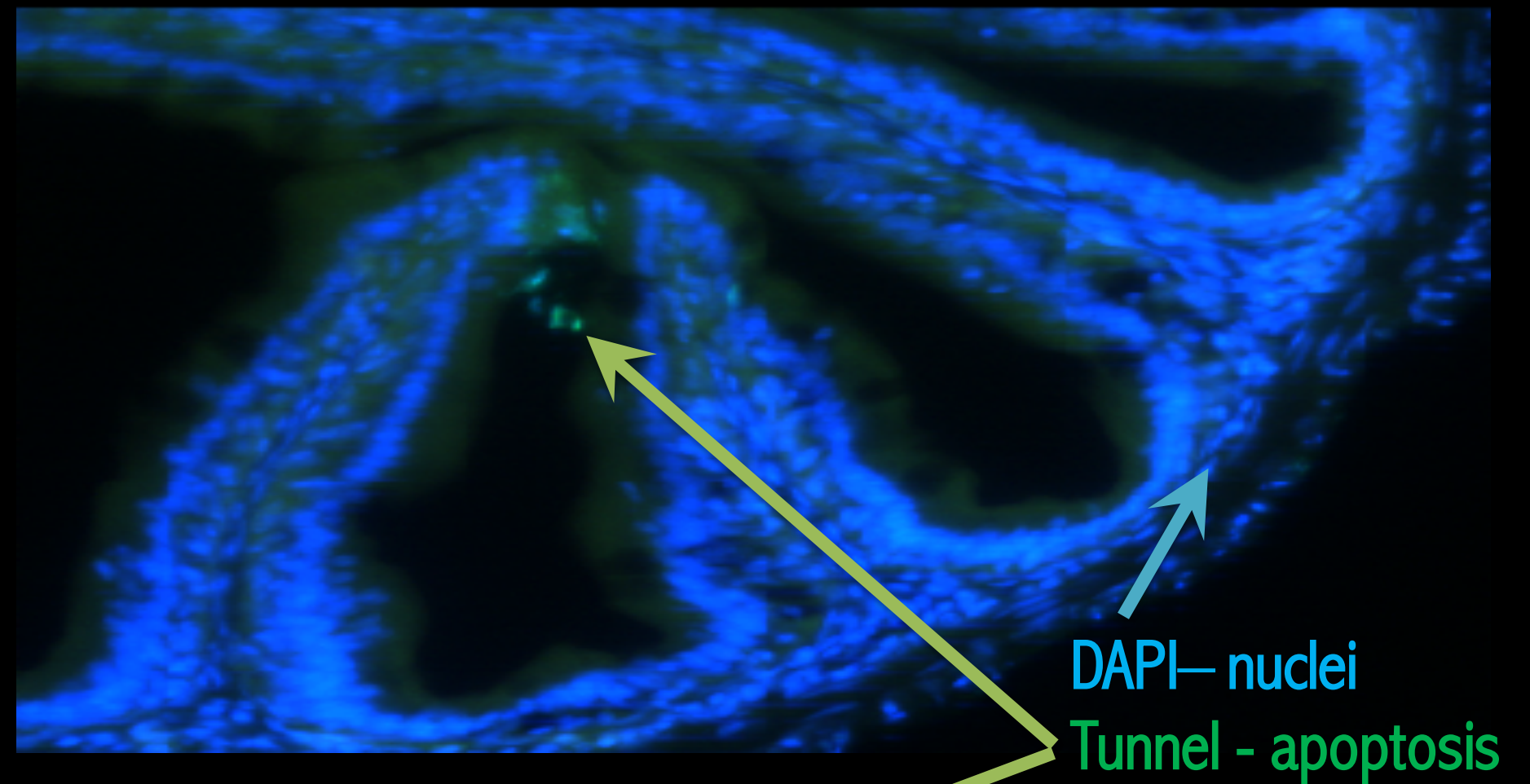


Fewer Apoptotic cells in Brooding  
Cichlids Compared to Fasted and Fed  
Female Cichlids:

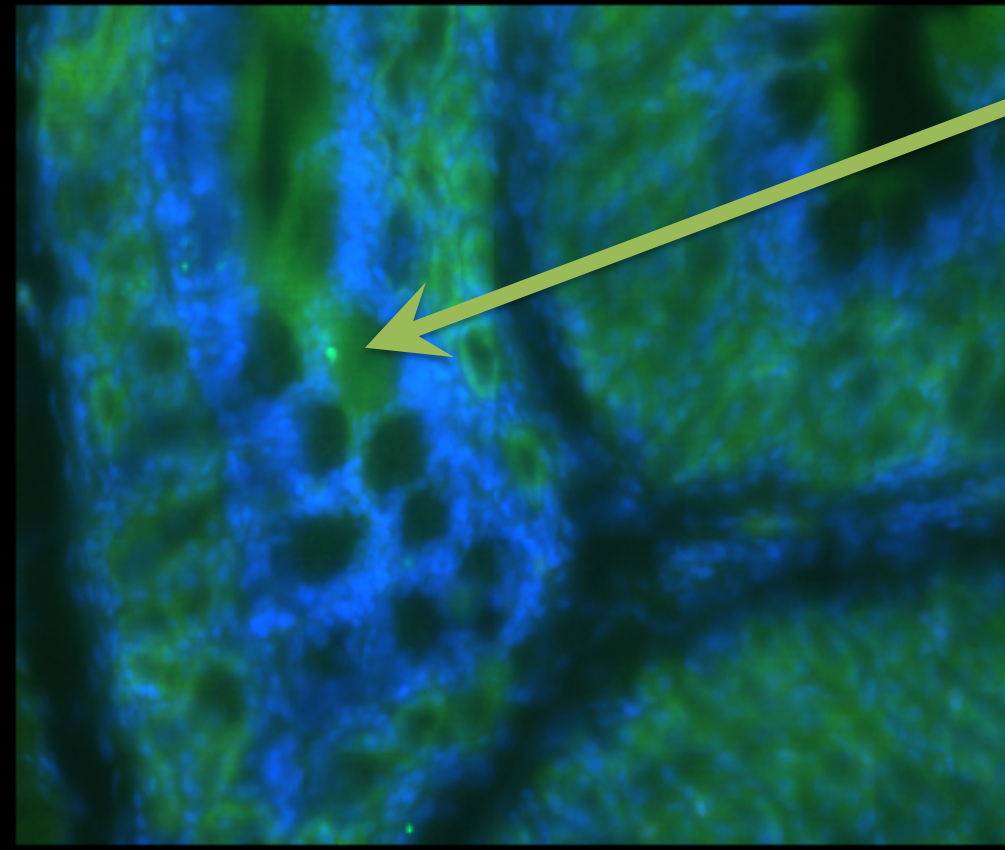




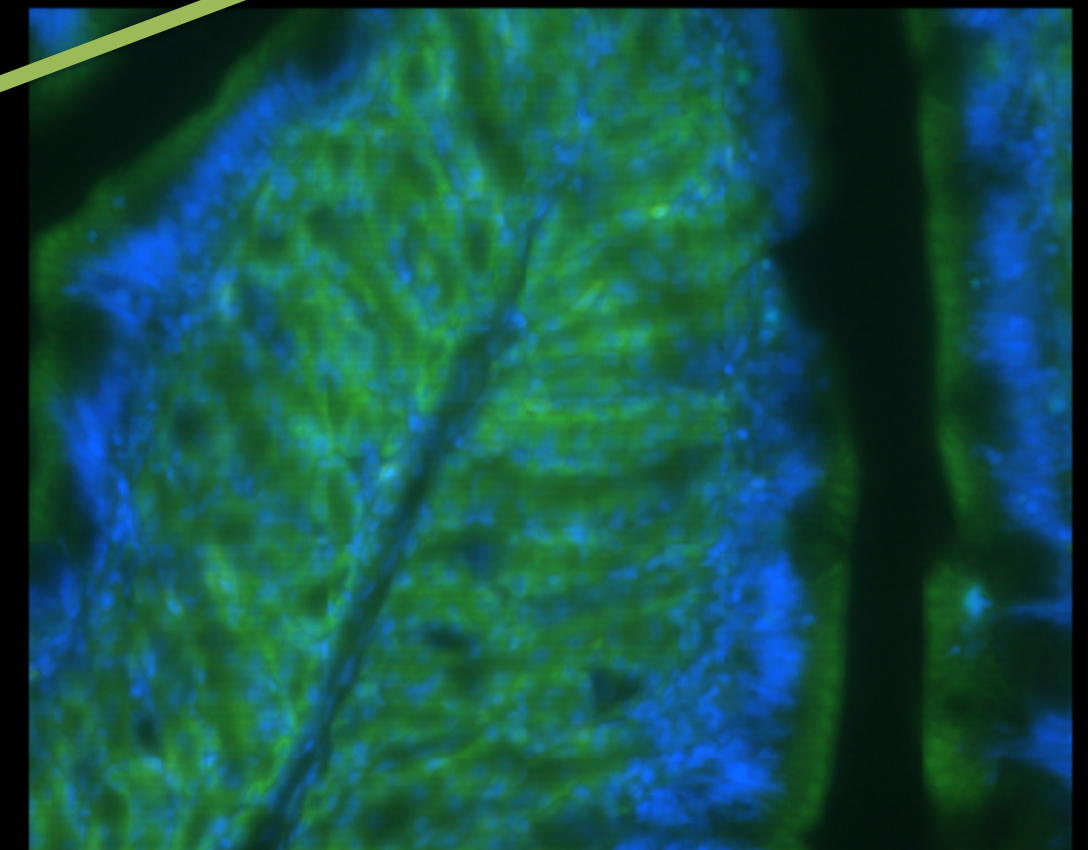
# TUNEL stained Apoptotic Cells in Intestinal Epithelium



Fed



Fasted



Brooding



# Data suggests that mouthbrooding females conserve energy by down regulating intestinal epithelial turnover

## Future Directions:

- **5-bromo-2'-deoxyuridine (BrdU)** will stain for proliferating cells
- Examine the difference in cell turnover throughout the brooding cycle and in different locations in the intestine
- Investigate peripheral adaptations and changes that occur during the brooding period



## References & Acknowledgements

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E.M. Chikwati, J. Gu, M.H. Penn, A.M. Bakke, A. Krogh "Intestinal epithelial cell proliferation and migration in Atlantic salmon, *Salmo salar* L.: effects of temperature and inflammation" *Cell Tissue Res* (2013) 353:123-137

N. Hemmer, D. Steinhagen, W. Drommer, W. Korting "Changes of intestinal epithelial structure and cell turnover in carp *Cyprinus carpio* infected with *Goussia carpelli* (Protozoa: Apicomplexa)" *Diseases of Aquatic Organisms* (1998) Vol. 34: 39-44

K P. Maruska, R.E. Carpenter, R.D. Fernald "Characterization of Cell Proliferation throughout the Brain of the African Cichlid Fish *Astatotilapia burtoni* and its Regulation by Social Status" *The Journal of Comparative Neurology* (2012) Vol: 520: 3471-3491

H. Takahashi, T. Sakamoto, K. Narita "Cell proliferation and apoptosis in the anterior intestine of an amphibious, euryhaline mudskipper (*Periophthalmus modestus*)" *J Comp Physiol B* (2006) 176: 463-468